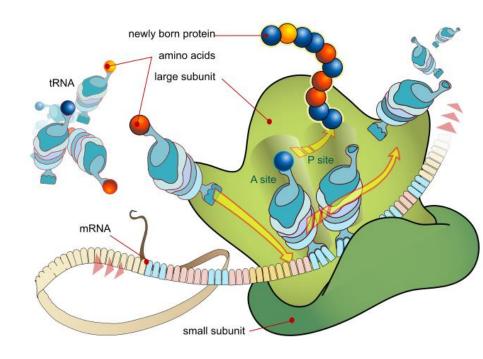
# **RNA MACHINERIES**



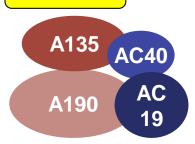
TRANSCRIPTION - RNAP+ SPLICING - SPLICEOSOME 3'end FORMATION - CPA



**TRANSLATION - RIBOSOME**<sup>+</sup>

# RNA POLYMERASES

Core subunits (similar in all)



RNA Pol I

Common subunits (same in all)

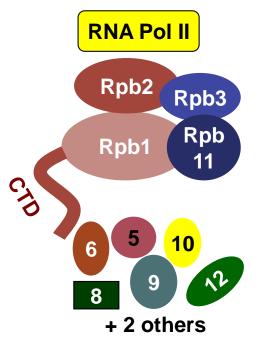


+ 4 others

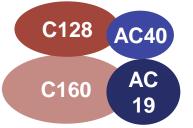
RNAs: ribosomal RNA

35S precursor contains 18S, 5.8S and 25S rRNAs

**Additional plant Polymerases** 



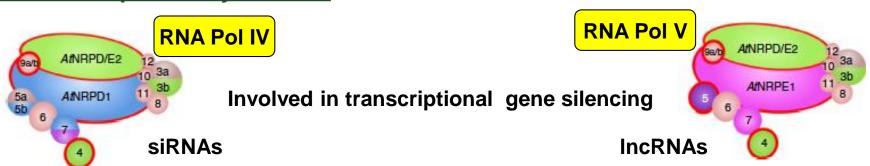
mRNA, most snRNAs (U1, U2, U3, U4, U5, U11, U12, U4atac), snoRNAs, microRNAs, telomerase RNA, ncRNAs RNA Pol III





+ 5 others

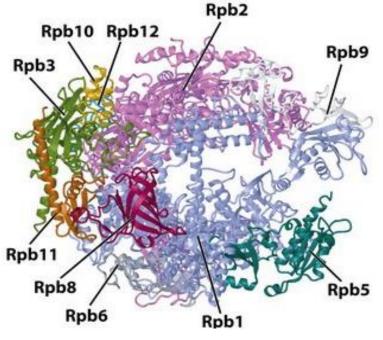
tRNA, 5S rRNA, U6 snRNA, U6atac snRNA, 7SK RNA, 7SL RNA, RNase P RNA, RNase MRP RNA



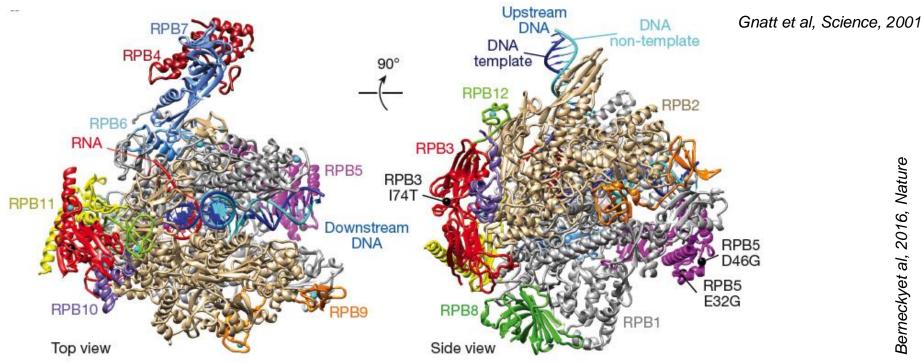
Zbigniew Dominski, lectures 2008

#### Yeast Pol II

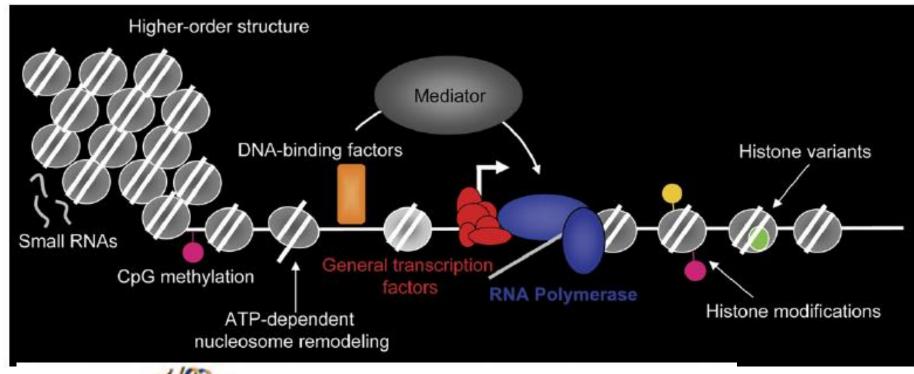
- 12 subunits
- core by specific Rpb1-3, 9 and 11
- Rpb5-6, 8, 10 and 12 shared by Pol I-III
- specific subcomplex Rpb4/7 not essential
- associated factors RAP74, RAP30 (TFIIF)

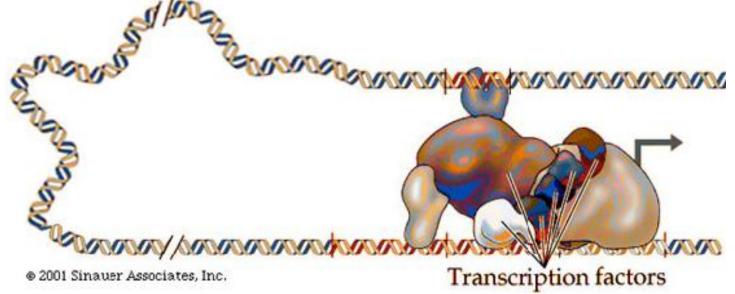


#### **Mammalian Pol II**

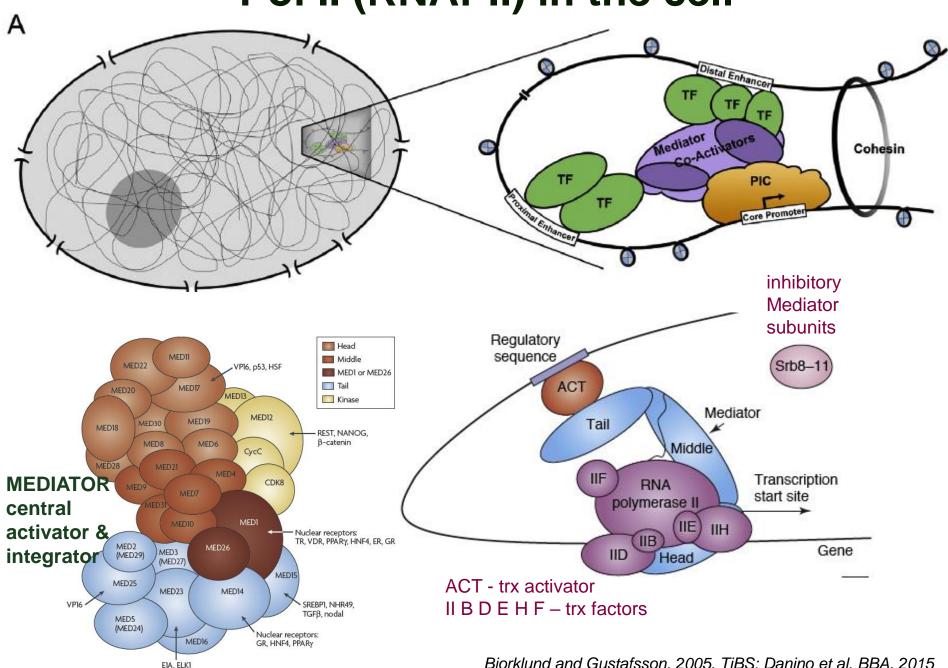


# Pol II (RNAPII) in the cell



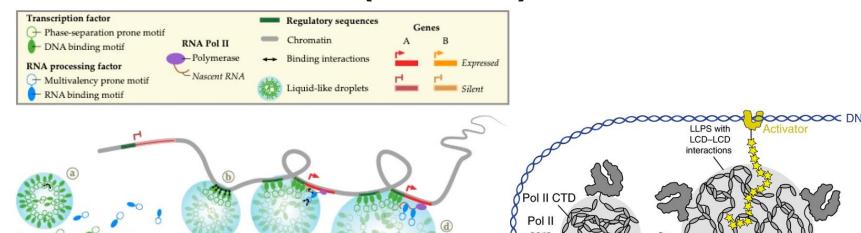


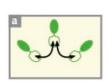
# Pol II (RNAPII) in the cell



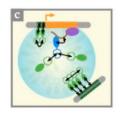
Bjorklund and Gustafsson, 2005, TiBS; Danino et al. BBA, 2015

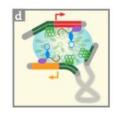
# Pol II (RNAPII) in the cell











#### LLPS, droplets

Liquid-like phase separation
Transcriptional condensates are formed
by phase-separation self-assembly driven
by IDR (Intrinsically Disordered Region)containing proteins (e.g. CTD in Pol II)

#### **CTD-driven phase separation**

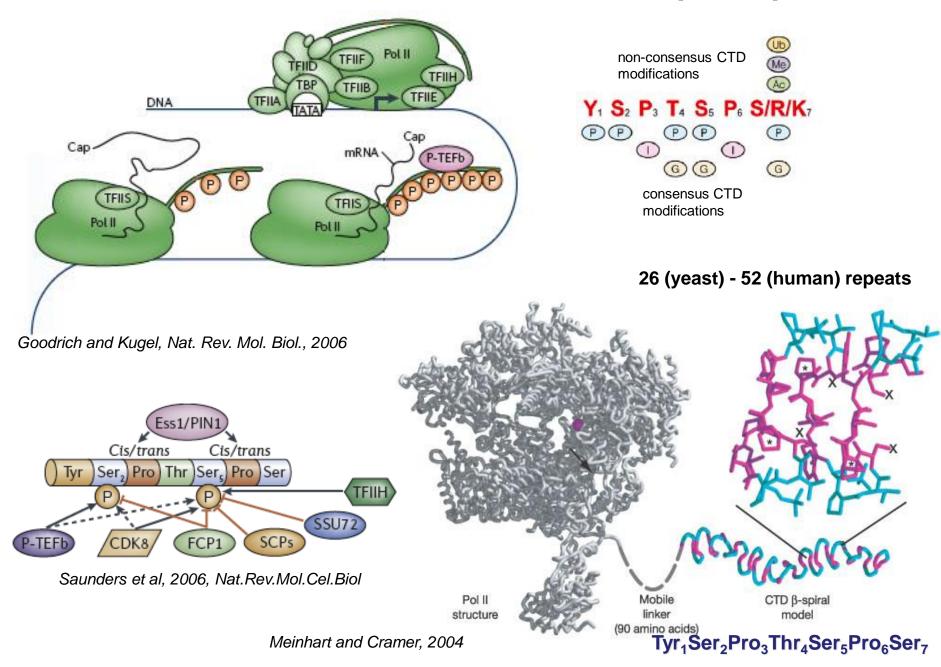
Activators recruit/nucleate Pol II hubs near promoters. Initiation-coupled CTD phosphorylation removes individual Pol II enzymes for transcription elongation.

Initiation

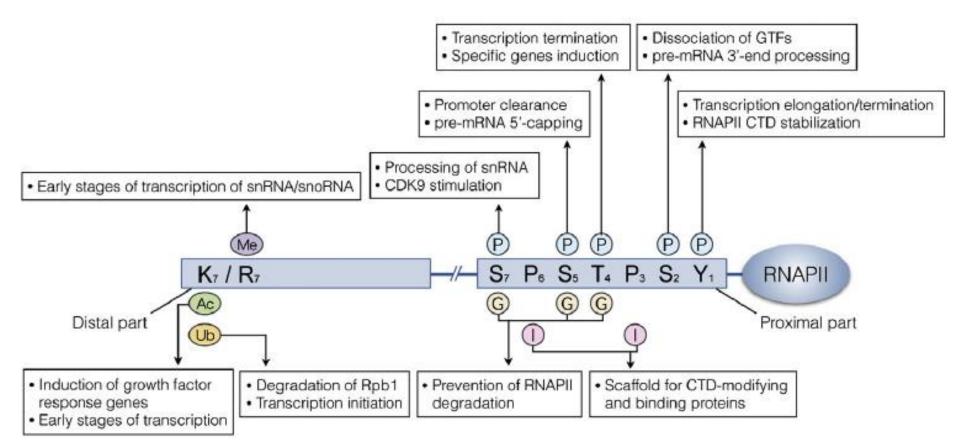
Lesne et al.,2019 Genes Boehning et al, 2018, Nat Struct Mol Biol

Elongation

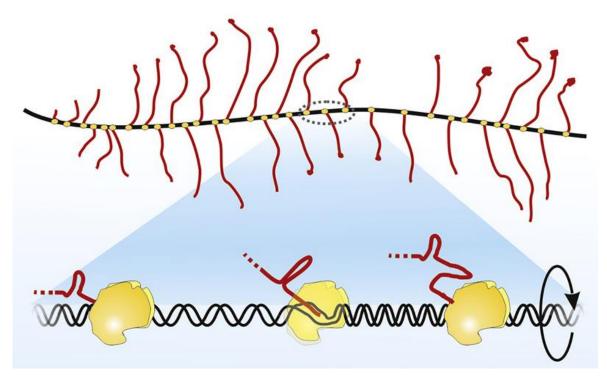
# Pol II C-terminal domain (CTD)



### CTD CODE



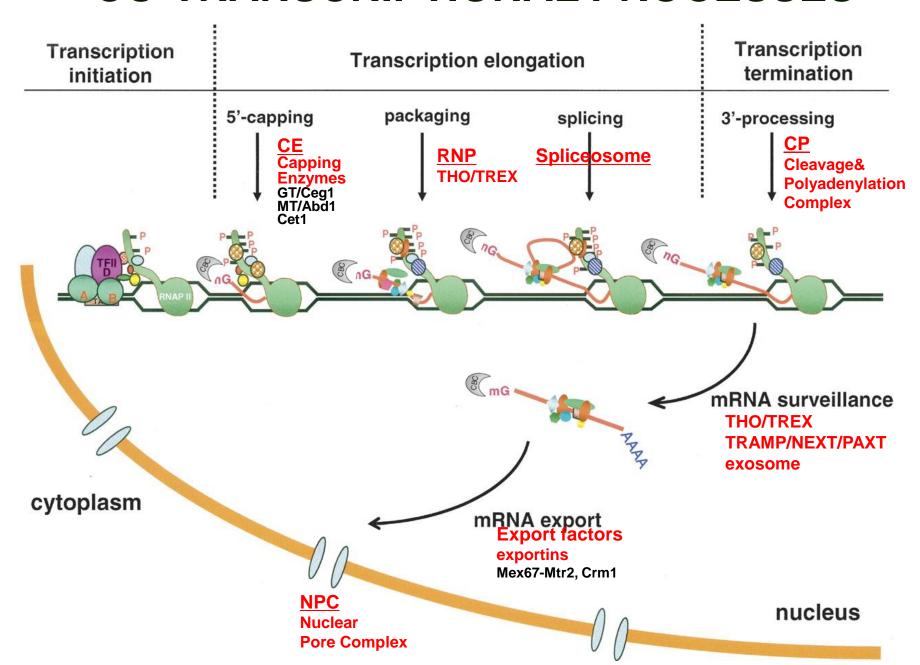
# **NASCENT TRANSCRIPTS**



#### Nascent transcript = during formation, newly formed, still bound by Pol II

- nascent RNAs couple RNA processing with transcription elongation and chromatin modification
- nascent RNAs modulate binding of proteins to regulatory elements (chromatin)
- regulatory effects of nascent transcripts can be enhanced by gene looping
- high concentrations of nascent RNAs can initiate formation of nuclear bodies
- sometimes the function is conferred by nascent transcription (activity) and not the transcript itself

# **CO-TRANSCRIPTIONAL PROCESSES**

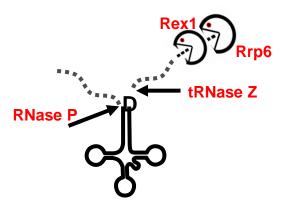


Li and Manley, Genes Dev, 2006

### POST-TRANSCRIPTIONAL PROCESSES

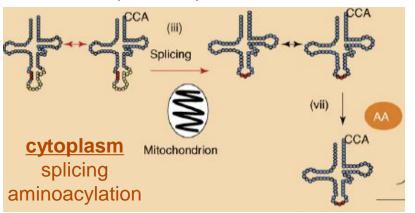
#### **tRNA PROCESSING**

- 5' end by RNAse P
- 3' end by tRNase Z or
- by exonuclease Rex1 and Rrp6

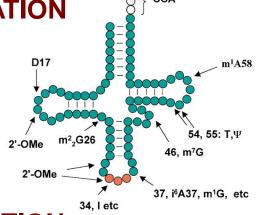


#### **tRNA SPLICING**

In the cytoplasm on the mitochondrial membrane (YEAST!!)



tRNA MODIFICATION
by RNA modifying
enzymes

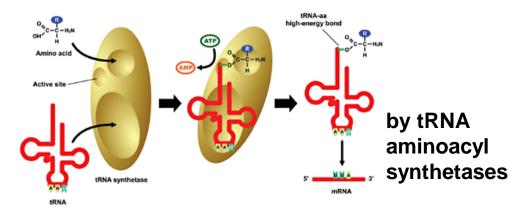


**tRNA CCA ADDITION** 

by tRNA nucleotidyltransferase



#### **tRNA AMINOACYLATION**



Hopper and Shaheen, TiBS,2008

PIC

**Preinitiation** 

**Complex** 

Scaffold

factors

transcription

(TFIID, A, E, H)

CF1, CPF

**Complex** 

# **GENE LOOPING**

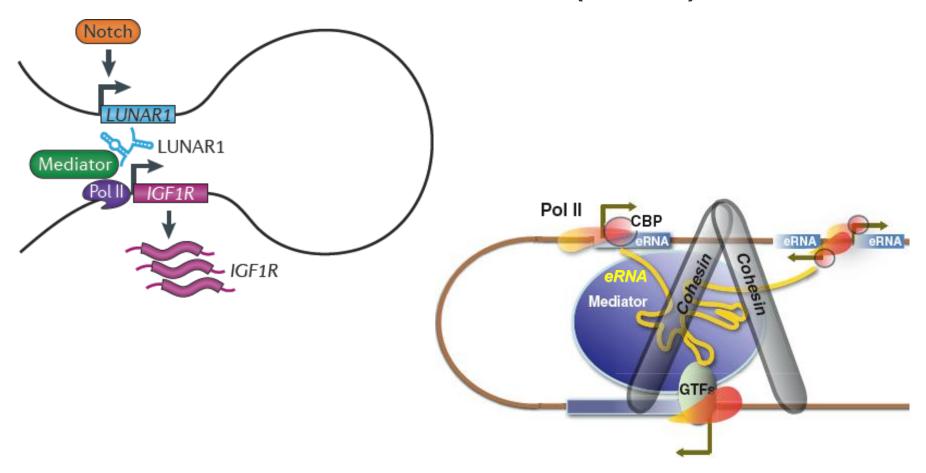
Pol II (also Pol I) Recruitment of PIC Terminator Promoter Initiation of Transcription Mediator Scaffold RNAP Reinitiation of Transcription Cleavage and **Polyadenylation** 

Loop formation requires interaction between factors at the promoter (THIIB) and terminator (Rna15 from CF1) /in mammals: transcription factors, nuclear receptors, insulators, chromatin remodellers, Polycomb, architectural proteins/

Loop function: facilitation of transcription reinitiation of PollI, but also repression of gene expression (PcG, DNA methylation)

### **GENE LOOPING**

via Mediator and enhancer RNAs (eRNAs)



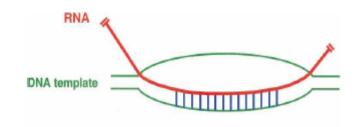
Some eRNAs (e.g. LUNAR1 near the IGF1R locus) mediate chromosome looping between enhancers and nearby genes via Mediator or MLL protein complexes

Quinn and Chang, Nat Rev

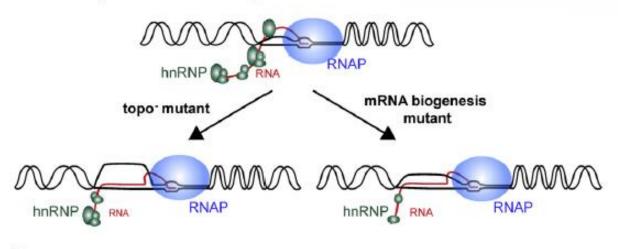
Quinn and Chang, Nat Rev Genet 2015; Lai and Shiekhattar, Curr Op Gene Dev 2014

### R-LOOPs

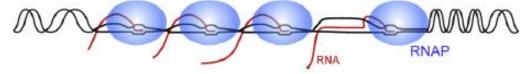
# DNA::RNA hybrids formed during transcription before RNP packaging



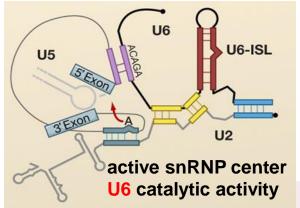
A Transcription associated R-loop formation



B RNAP roadblock



- R-loops
- accumulate in RNP biogenesis mutants (tho, sen1, mRNA export)
- negative effects: polymerase stalling, termination defects, replication fork stalling, DNA damage, genetic instability
- prevented by topoisomerases, helicase Sen1, THO complex, resolution (cleavage) by RNase H



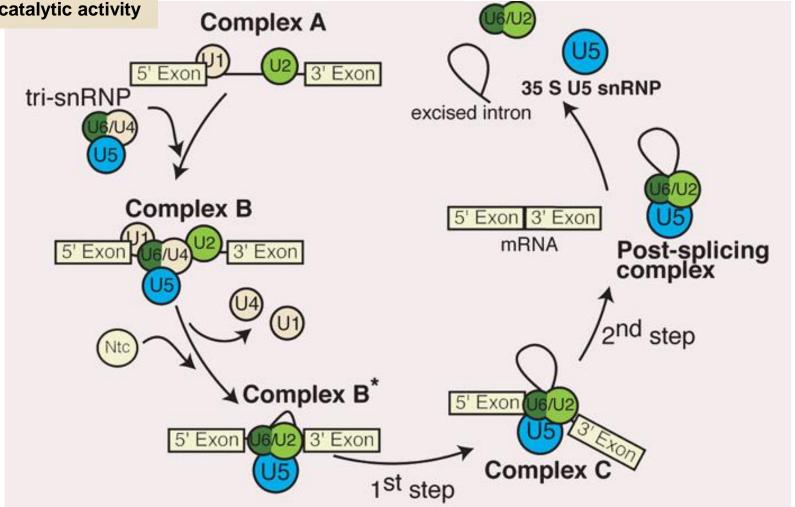
# **SPLICEOSOME**

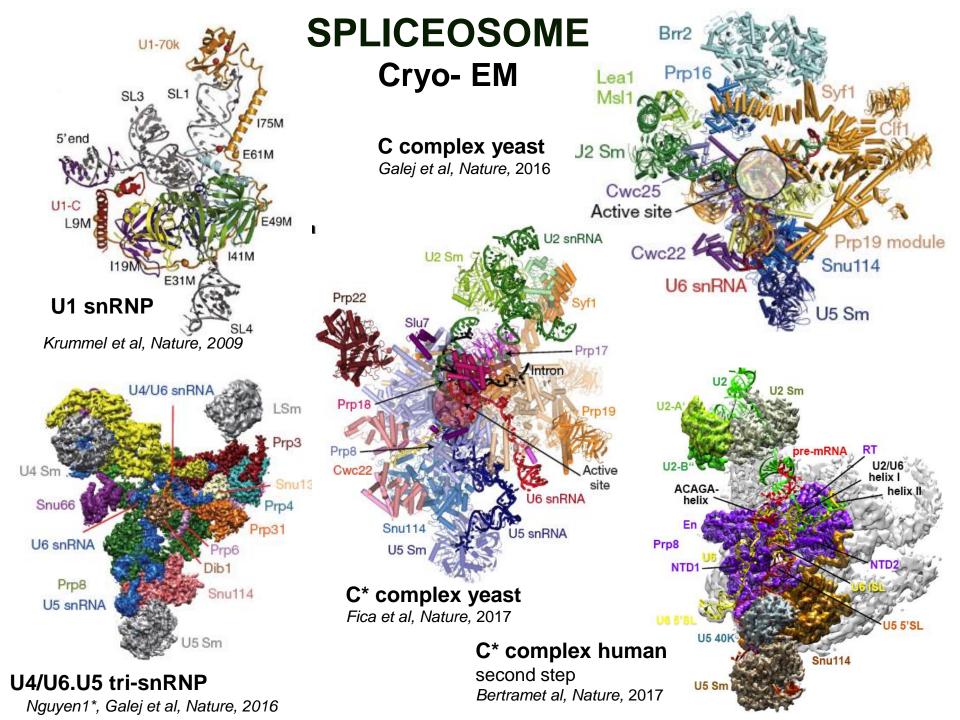
5 snRNAs: U1, U2, U4, U5, U6

Core Sm or LSM (U6) proteins

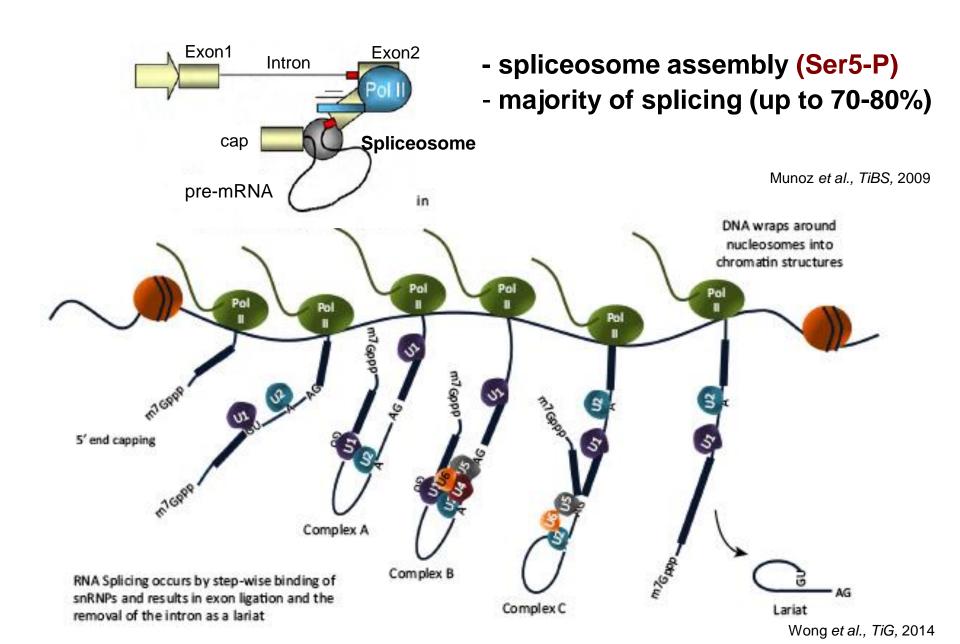
1.7 - 3 MDa Specific snRNP proteins

**Splicing factors** 

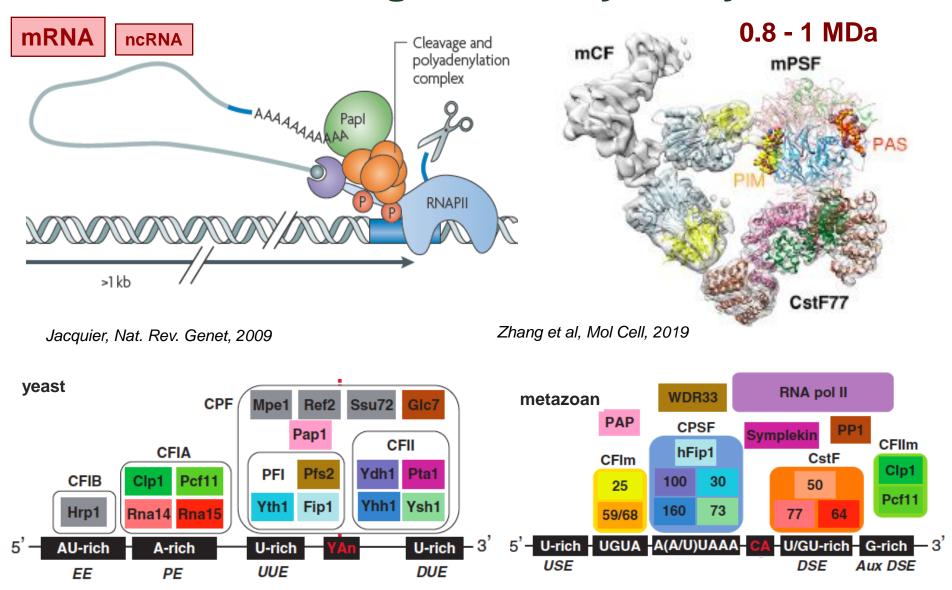




# SPLICING: co-transcriptional process

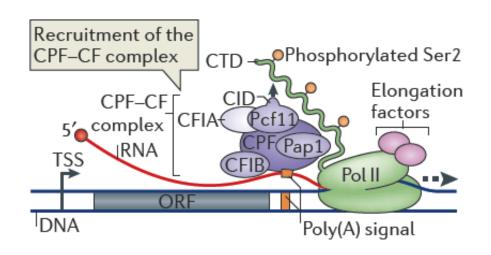


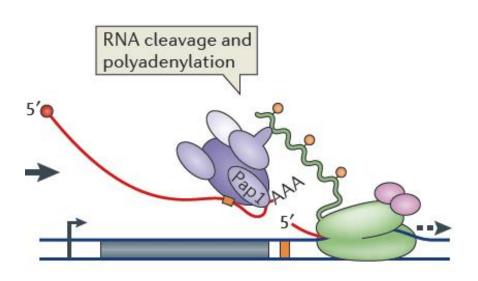
# **CPA Cleavage and Polyadenylation**

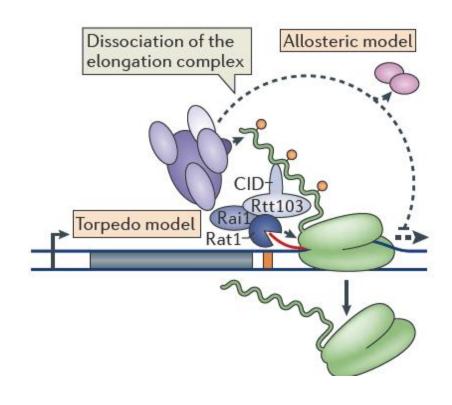


Cleavage by CPSF-73 (human), Brr5/Ysh1 (yeast)

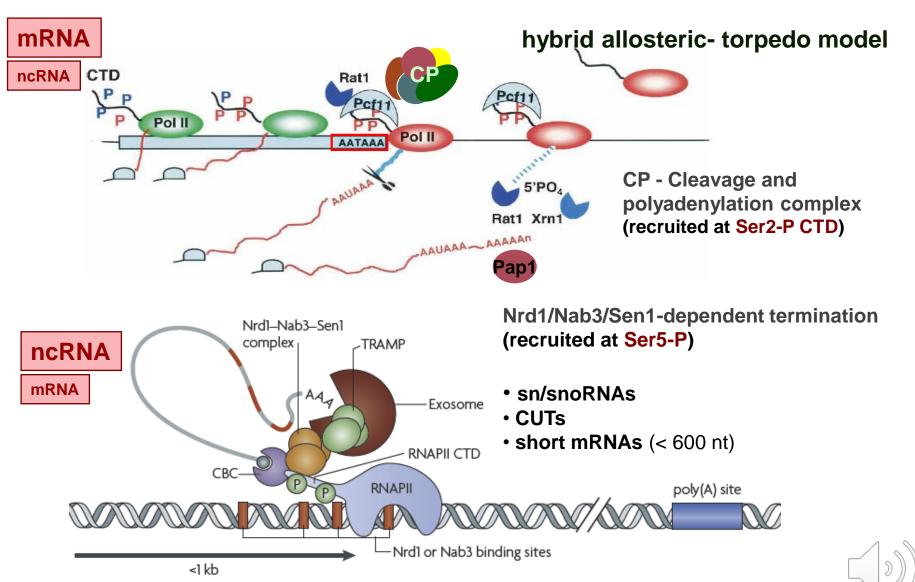
# CPA: mRNA 3' end formation transcription termination at mRNA genes



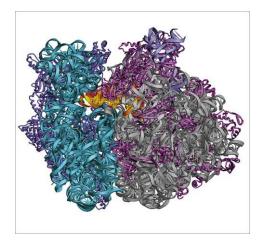




# POL II TRANSCRIPTION TERMINATION



# **RIBOSOME**



3.3 (yeast) - 4.3 (humans) MDa

### Ribosome is a ribozyme

- No ribosomal protein with a peptidyl transferase (PT) activity
- Drugs (chloramphenicol) that inhibit PT bind to the 25S rRNA (PT loop)
- Mutations that provide resistance to these drugs map to the PT loop
- Nearly all (99%) of proteins can be stripped from the large subunit and it still retains the PT activity
- Only RNA chains are close enough to the PT center (structure)
- Ribosomal proteins are important for ribosome stability and integrity, but NOT for catalysis

# TRANSLATION CYCLE

